

Research Article

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Prevalence and intensity of Streblidae in bats from a Neotropical savanna region in Brazil

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Abstract: Bats of the family Phyllostomidae are common hosts to streblids known as bat flies. Here, we discuss the component community, prevalence and intensity of infection with species of Streblidae on an assemblage of phyllostomid bats in the Cafuringa Environmental Protection Area (APA Cafuringa) in the core area of the Cerrado in Central Brazil. A total of 1 841 streblid individuals of 24 species occurred on 752 bats of 14 species. Ten species of streblids infected *Glossophaga soricina* (Pallas), whereas seven or fewer streblid species infected the other bat species. Nine bat fly species presented a prevalence of more than 50%, whereas some differences in the abundance of bat flies among hosts were observed. *Strebla wiedemanni* Kolenati, 1856 and *Trichobius furmani* Wenzel, 1966 were more host-specific compared to the other streblids, and they occurred in greater abundance on their preferred hosts. *Trichobius uniformis* Curran, 1935 and *Strebla mirabilis* (Waterhouse, 1879) were the least host-specific, occurring on five and six hosts, respectively.

Keywords: Diptera, ectoparasites, Chiroptera, Cerrado, South America

Bat flies, i.e. species of the family Streblidae (Diptera: Hippoboscoidea) are obligate ectoparasites of Chiroptera (Dick and Patterson 2006) and are typically associated with bats of the family of Phyllostomidae (Dittmar et al. 2006). Phyllostomid bats are unique to the New World, with the majority of species found in tropical and sub-tropical regions (Whitaker 1988). Streblids may possess normal, reduced or no wings. Their eyes may be small or absent and their bodies may be laterally or dorsoventrally flattened, with legs that can be short and robust or long and thin (Whitaker 1988).

Azevedo and Linardi (2002) reported that few data about streblid flies are available in Brazil. Several studies have been published but mainly from the Atlantic Forest (França et al. 2013), which is one of the five Brazilian biomes. As noted by Bertola et al. (2005), several studies have been conducted on the taxonomy of Streblidae, but only a few explored host-parasite relationships.

Seventy species belonging to 20 genera of streblid bat flies are registered in Brazil (Graciolli et al. 2010, Graciolli and Azevedo 2011). However, these numbers certainly underestimate the real richness of bat flies (Dias et al. 2009,

Graciolli et al. 2010). In fact, more than 90% of the vast Brazilian territory has not been properly surveyed for bats (Bernard et al. 2011). Consequently, reports on their ectoparasites are even more rare, and if the real richness of bat flies is unknown, then the relationship between bat flies and their hosts is probably in the same category.

The parasitism of streblids may be affected by some factors intrinsic to the bat hosts, such as body size, grooming behaviour and immunological defences (Ter Hofstede and Fenton 2005, Patterson et al. 2007, 2008). Furthermore, the different types of bat roosts and the sharing of roosts among individual bats and among different bat species are known to affect the distribution and abundance of streblids on the host populations (Patterson et al. 2007). From another point of view, the knowledge of bat ectoparasites provides valuable information about the biology, systematics, and phylogeny of the host (Fritz 1983).

Host specificity is typical for parasite-host associations. Obligate or permanently present parasite associations are usually higher (Dick et al. 2009) and, therefore, ectoparasites can be classified as heteroxenous, which is when they can infect many species in many genera. The term mon-

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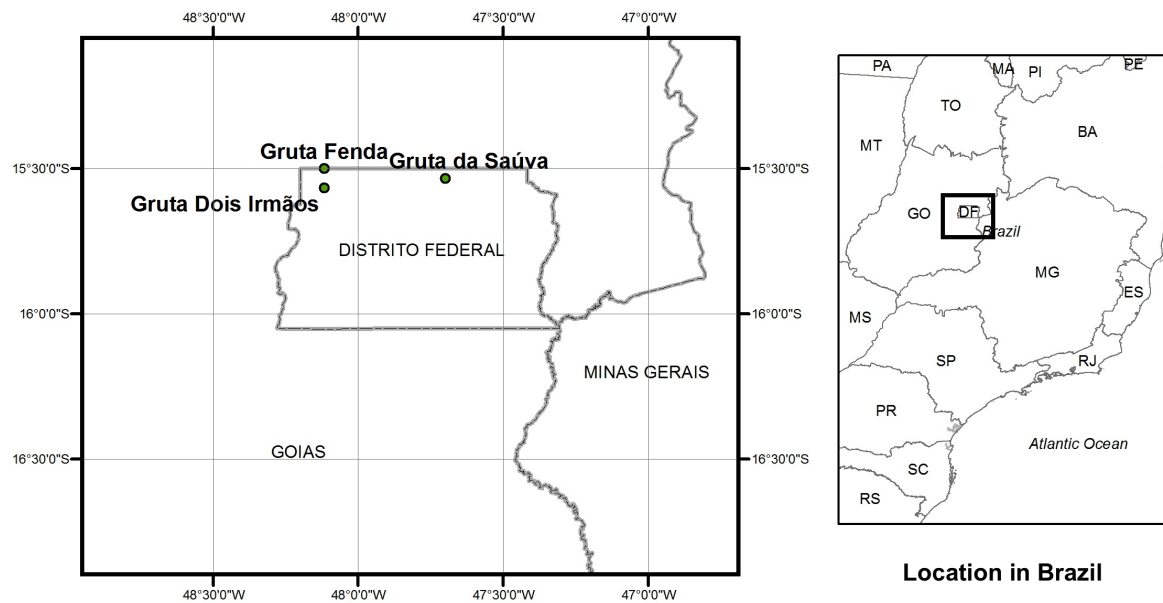


Fig. 1. Map of the caves of the Cafuringa Environmental Protection Area in the Cerrado of the Federal District, Brazil.

oxenous describes ectoparasites found on only one host species. When the ectoparasites are on two or more host species of the same genus, they are stenoxenous, and when they are regularly established on hosts belonging to different genera, they are oligoxenous (Ter Hofstede et al. 2004).

For the Cerrado region of Brazil, there are only five studies that report quantitative parasitological indices for the streblid communities (Coimbra et al. 1984, Komeno and Linhares 1999, Aguiar and Antonini 2011, Eriksson et al. 2011, Santos et al. 2013). Herein, we present a study about the component community, prevalence and the intensity of a infection of Streblidae on a bat assemblage (Phyllostomidae) present in the Cafuringa Environmental Protection Area (APA Cafuringa) in the core area of the Cerrado biome, a Neotropical savanna. The purpose of this paper was to study host-parasite relationships, evaluating the degree of variability in the streblid distribution in the different host populations.

MATERIALS AND METHODS

Study area

We captured bats monthly from April 2004 to March 2005 in three caves with 90 to 773 metres of horizontal development in the interfluvial mesophytic forests within the Cafuringa Environmental Protection Area (CEPA). The CEPA is located northwest of Brasília (15°30'–15°40'S; 47°50'–48°12'W) and is a 46 000 ha area characterised by karst geomorphology with steep and dry relief (Fig. 1). The climate of the region is characteristic of the Cerrado biome, with a dry winter and a rainy summer, and it is classified as Aw Köppen (Alvares et al. 2013).

Collection and identification of ectoparasites

Streblids flies were collected from bats captured with mist nets placed near the entrance of the following three caves (Fig. 1): Saúva is a dry limestone cave (15°32'S; 47°42'W) with 235 m of development, having three parallel segments connected by secondary conduits. It is poorly ventilated and can be accessed only

by a small entrance. The external vegetation is mainly gallery forest. Dois Irmãos is a dry limestone cave (15°34'S; 48°07'W) with 90 m of development, consisting of several small rooms at two superimposed levels and one entry. It is placed on a limestone hill covered by a mesophytic interfluvial forest and surrounded by savanna and crops. Fenda is the largest granite cave (15°30'S; 48°10'W) in the Distrito Federal, located at 840 m above sea level. It has 341 m of development, with a number of smaller channels and two large entrances. Since it is open to visitation, this cave is constantly disturbed. The original cerrado vegetation that surrounds the three caves has been heavily deforested and replaced by pastures or degraded semi-deciduous forests (Bredt et al. 1999, Aguiar et al. 2014).

One or two nylon mist nets (6 × 3 m) were used from 6 PM to 6 AM to capture the bats. After their capture, bats were marked, weighed and measured, their ectoparasites were removed, and then they were released. Nets were continuously checked to avoid the presence of too many bats at the same time in the net. We usually had one or two bats at the same time, but we avoided contact between the bats and the exchange of flies by removing the bats quickly for processing. Each bat removed from the mist net was kept in a separate cloth bag until processing to prevent the ectoparasites from escaping and to avoid sample contamination. All ectoparasites observed on the bats were removed with forceps and preserved in 70% ethanol. The species of bat flies were identified using the keys for Neotropical bat flies (Wenzel 1976, Guerrero 1993, 1994a,b, 1995a,b, 1996, 1998). Identification was confirmed by G. Gracioli and all vouchers of the ectoparasites species were deposited in the Chiroptera Collection at the University of Brasília (CCUNB).

Data analysis

The definitions of component community (the assemblage of parasite species on a given bat species), prevalence (the percentage of bats infected by a given parasite species) and the intensity of infection (the number of flies of a given species of bat) followed Bush et al. (1997). Confidence intervals were calculated

Table 1. Number of Streblidae and bat species (N), number of infected hosts individuals (IH), prevalence (P) and mean intensity of infection (MI) with bat flies collected on phyllostomid bats in the Cafuringa Environmental Protection Area in the Cerrado of Central Brazil. * accidental

Host (N) and their streblid species	Streblidae (N)	IH	P (%)	MI
<i>Anoura caudifer</i> (É. Geoffroy Saint-Hilaire) (3)				
<i>Anastrebla caudiferae</i> Wenzel, 1976	1	1	33	1.0
<i>Anastrebla modestini</i> Wenzel, 1966	1	1	33	1.0
<i>Strebla harderi</i> Wenzel, 1976	1	1	33	1.0
<i>Trichobius propinquus</i> Wenzel, 1976	1	1	33	1.0
<i>Trichobius tiptoni</i> Wenzel, 1976	2	1	33	1.0
<i>Carollia perspicillata</i> (Linnaeus) (32)				
<i>Paraeuctenodes longipes</i> * Pessoa et Guimaraes, 1937	1	1	3	1.0
<i>Speiseria ambigua</i> Kessel, 1925	3	2	6	1.0
<i>Strebla guajiro</i> (Garcia et Casal, 1965)	12	10	31	1.2
<i>Trichobius dugesii</i> Townsend, 1891	2	2	6	1.0
<i>Trichobius joblingi</i> Wenzel, 1966	93	28	91	3.3
<i>Trichobius tiptoni</i>	3	1	3	3.0
<i>Trichobius uniformis</i> Curran, 1935	3	3	6	1.0
<i>Chrotopterus auritus</i> (W. Peters) (4)				
<i>Strebla chrotopteri</i> Wenzel, 1976	11	4	75	3.7
<i>Strebla mirabilis</i> (Waterhouse, 1879)	5	1	25	5.0
<i>Trichobius longipes</i> (Rudow, 1871)	6	1	25	2.0
<i>Desmodus rotundus</i> (E. Geoffrey) (346)				
<i>Strebla mirabilis</i>	1	1	0.5	1.0
<i>Strebla wiedemanni</i> Kolenati, 1856	540	151	44	3.6
<i>Trichobius furmani</i> Wenzel, 1966	57	37	11	1.5
<i>Trichobius parasiticus</i> Gervais, 1844	214	102	30	2.1
<i>Diaemus youngi</i> (Jentik) (8)				
<i>Strebla diaemi</i> Wenzel, 1966	51	8	100	6.4
<i>Trichobius diaemi</i> Wenzel, 1976	84	8	100	10.5
<i>Diphylla ecaudata</i> Spix (265)				
<i>Strebla mirabilis</i>	6	5	2	1.2
<i>Strebla wiedemanni</i> *	4	3	1	1.3
<i>Trichobius furmani</i>	372	114	43	3.3
<i>Trichobius parasiticus</i>	9	4	2	2.2
<i>Trichobius uniformis</i>	5	1	0.4	5.0
<i>Glossophaga soricina</i> (Pallas) (29)				
<i>Paraeuctenodes longipes</i>	10	5	17	2.0
<i>Paratrachobius longicrus</i> (Miranda-Ribeiro, 1907)	3	2	7	1.5
<i>Speiseria ambigua</i>	3	3	10	1.0
<i>Strebla altmani</i> Wenzel, 1966	2	1	6	1.0
<i>Strebla guajiro</i>	5	5	17	1.0
<i>Strebla mirabilis</i> *	1	1	3	1.0
<i>Trichobius dugesii</i>	13	9	31	1.4
<i>Trichobius furmani</i>	1	1	3	1.0
<i>Trichobius joblingi</i>	11	8	24	1.7
<i>Trichobius uniformis</i>	10	7	31	1.1
<i>Lonchophylla dekeyseri</i> Taddei, Vizotto et Sazima (38)				
<i>Speiseria ambigua</i>	11	8	21	1.2
<i>Strebla altmani</i>	13	5	13	1.4
<i>Strebla guajiro</i>	8	6	16	1.3
<i>Trichobius dugesii</i>	4	1	3	4.0
<i>Trichobius uniformis</i>	118	31	81	3.6
<i>Lonchorhina aurita</i> Tomes (3)				
<i>Trichobius dugesii</i>	1	1	33	1.0
<i>Trichobius dugesioides</i> Wenzel, 1966	18	3	100	6.0
<i>Micronycteris minuta</i> (Gervais) (1)				
<i>Trichobius joblingi</i>	3	1	100	3.0
<i>Phyllostomus hastatus</i> (Pallas) (12)				
<i>Strebla mirabilis</i>	10	4	33	2.5
<i>Trichobius longipes</i>	32	9	83	3.2
<i>Platyrrhinus lineatus</i> (E. Geoffroy) (3)				
<i>Neotrichobius delicatus</i> (Machado-Allison, 1966)	1	1	33	1.0
<i>Paratrachobius longicrus</i>	4	2	67	1.5
<i>Sturnira lilium</i> (E. Geoffroy) (1)				
<i>Megistopoda proxima</i> (Seguy, 1926)	5	1	100	2.0
<i>Trachops cirrhosus</i> (Spix) (7)				
<i>Trichobius dugesioides</i>	66	7	100	1.1
Total = 752	1 841			

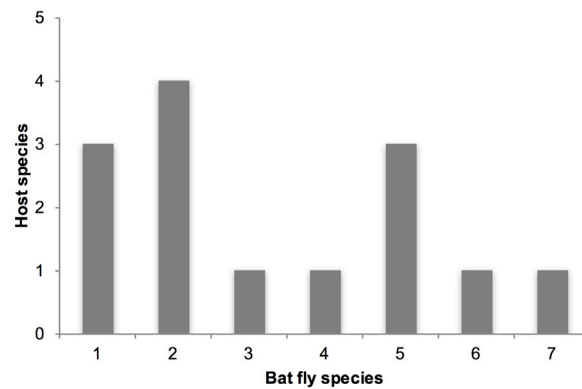


Fig. 2. Relation of the number of bat fly species on infected hosts, captured from April 2004 to March 2005 at the caves of Cafuringa Environmental Protection Area in the Federal District, Brazil.

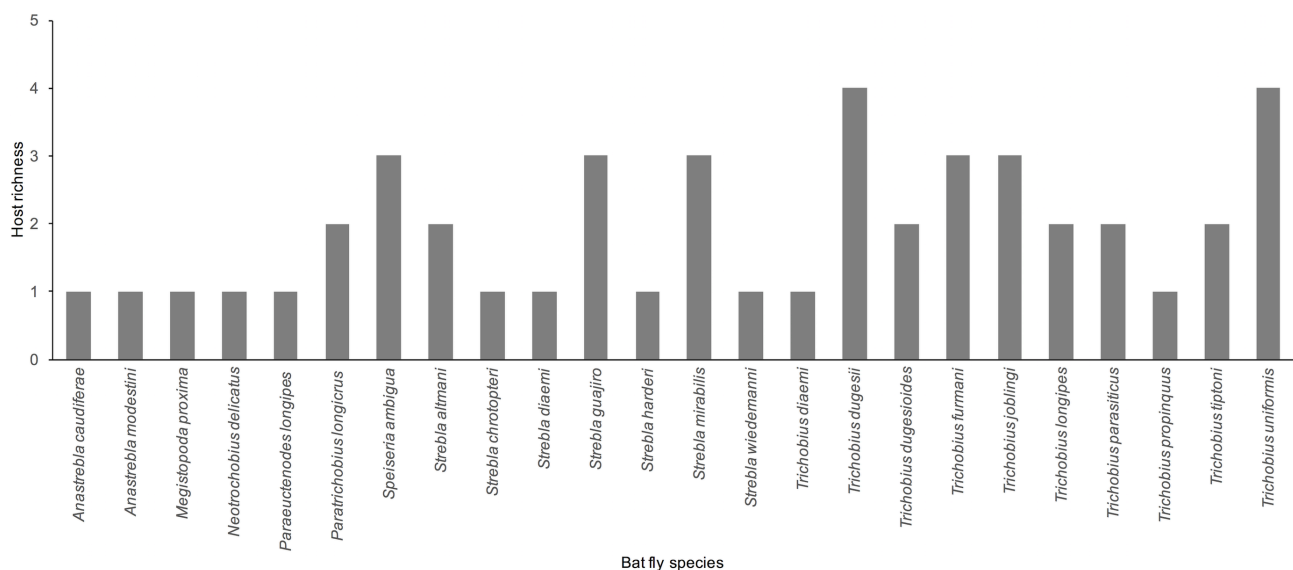


Fig. 3. Host richness of bat fly species on hosts captured from April 2004 to March 2005 at the caves of the Cafuringa Environmental Protection Area in the Federal District, Brazil (accidental records are not considered)..

using Quantitative Parasitology 3.0 (Rózsa et al. 2000). Non-primary infections (Dick 2007) were not considered to enable comparisons with other studies. Mean abundance refers to the number of ectoparasites per number of examined bats. Mean intensity refers to the average number of ectoparasites per number of infected bats. Host specificity was calculated using the Specificity Index (SI) (Marshall 1982). Since the data for ectoparasites had a non-normal distribution, we used a non-parametric analysis of variance (Kruskal-Wallis test) to gauge the preference of specific ectoparasites for a given species of bat host. The type of bat host was the categorical variable and the number of ectoparasites of each species was the continuous variable.

RESULTS

Component community

We captured 892 individuals bats from 15 species of Phyllostomidae and 752 of them (i.e. 84% of the captured bats) were infected with streblid ectoparasites. Only one bat species, *Artibeus planirostris* (Spix), was free of bat flies. The other bat species were parasitised by 24 species

of streblids (n = 1 841 individuals) (Table 1). Among the captured bat flies, 11 (46%) were monoxenous and 13 (54%) were oligoxenous.

Infracommunity

Half of the bat species was infected by one or two bat fly species (Fig. 2). The species-richest infracommunities (all the ectoparasite species on an individual host, including accidental records) were on *Glossophaga soricina* (Pallas) (species = 10). *Micronycteris minuta* (Gervais), *Sturnira lilium* (E. Geoffroy) and *Trachops cirrhosus* (Spix) were each infected with only one streblid species; *Diaemus youngi* (Jentik), *Lonchorhina aurita* Tomes, *Phyllostomus hastatus* (Pallas) and *Platyrrhinus lineatus* (E. Geoffroy) were infected with two streblid species; *Chrotopterus auritus* (W. Peters) was infected with three species; *Desmodus rotundus* (E. Geoffroy) was infected with four streblid species and it was the most abundant bat species in this study site; *Anoura caudifer* (É. Geoffroy Saint-Hilaire), *Diphylla ecaudata* Spix and *Lonchophylla dekeyseri* Taddei, Vizotto et Sazima were infected with five streblid species each; and

Table 2. Number of bats of the Phyllostomidae family, their infracommunities of ectoparasitic flies (N), and percentage of bats parasitized in the CEPA Cafuringa in the Cerrado of Central Brazil.

Hosts (n)/Ectoparasite	N	%
<i>Anoura caudifer</i> (É. Geoffroy Saint-Hilaire) (2)		
<i>Anastrebla caudiferae</i> + <i>Trichobius tiptoni</i> Wenzel, 1976	1	50
<i>Anastrebla modestini</i> Wenzel, 1966 + <i>Trichobius propinquus</i> Wenzel, 1976	1	50
<i>Carollia perspicillata</i> (13)		
<i>Pareuctenoides longipes</i> Pessoa et Guimaraes, 1937 + <i>Trichobius dugesii</i> Townsend, 1891 + <i>Trichobius joblingi</i> + <i>Trichobius uniformis</i>	1	8
<i>Speiseria ambigua</i> Kessel, 1925 + <i>Trichobius uniformis</i> Curran, 1935	2	15
<i>Strebla guajiro</i> (García et Casal, 1965) + <i>Trichobius joblingi</i> Wenzel, 1966	9	70
<i>Trichobius dugesii</i> + <i>Trichobius joblingi</i>	1	8
<i>Chrotopterus auritus</i> (W. Peters) (1)		
<i>Strebla chrotopteri</i> Wenzel, 1976 + <i>Strebla mirabilis</i> (Waterhouse, 1879) + <i>Trichobius longipes</i> (Rudow, 1871)	1	100
<i>Desmodus rotundus</i> (E. Geoffroy) (32)		
<i>Strebla wiedemanni</i> Kolenati, 1856 + <i>Trichobius parasiticus</i> Gervais, 1844		
<i>Diaemus youngi</i> (Jentik) (8)		
<i>Strebla diaemi</i> Wenzel, 1966 + <i>Trichobius diaemi</i> Wenzel, 1976	8	100
<i>Dyphyla ecaudata</i> Spix (4)		
<i>Strebla mirabilis</i> + <i>Trichobius furmani</i> Wenzel, 1966	2	50
<i>Trichobius furmani</i> + <i>Trichobius parasiticus</i>	2	50
<i>Glossophaga soricina</i> (Pallas) (9)		
<i>Paraeuctenoides longipes</i> + <i>Strebla guajiro</i> + <i>Trichobius dugesii</i>	1	11
<i>Paraeuctenoides longipes</i> + <i>Trichobius dugesii</i>	1	11
<i>Paraeuctenoides longipes</i> + <i>Trichobius dugesii</i> + <i>Trichobius uniformis</i>	1	11
<i>Paraeuctenoides longipes</i> + <i>Trichobius joblingi</i>	2	22
<i>Speiseria ambigua</i> + <i>Strebla guajiro</i>	1	11
<i>Strebla guajiro</i> + <i>Trichobius dugesii</i> + <i>Trichobius uniformis</i>	1	11
<i>Strebla guajiro</i> + <i>Trichobius joblingi</i>	1	11
<i>Strebla guajiro</i> + <i>Trichobius joblingi</i> + <i>Trichobius uniformis</i>	1	11
<i>Lonchophylla dekeyseri</i> (15)		
<i>Strebla altmani</i> + <i>Speiseria ambigua</i> + <i>Trichobius uniformis</i>	1	7
<i>Strebla altmani</i> + <i>Trichobius dugesii</i> + <i>Trichobius uniformis</i>	1	7
<i>Strebla altmani</i> Wenzel, 1966 + <i>Trichobius uniformis</i>	4	27
<i>Speiseria ambigua</i> + <i>Strebla guajiro</i>	1	7
<i>Speiseria ambigua</i> + <i>Strebla altmani</i> + <i>Strebla guajiro</i> + <i>Trichobius uniformis</i>	1	7
<i>Speiseria ambigua</i> + <i>Trichobius uniformis</i>	4	27
<i>Strebla guajiro</i> + <i>Trichobius uniformis</i>	3	20
<i>Lonchorhina aurita</i> Tomes, 1863 (1)		
<i>Trichobius dugesii</i> + <i>Trichobius dugesioides</i> Wenzel, 1966	1	100
<i>Phyllostomus hastatus</i> (Pallas) (1)		
<i>Strebla mirabilis</i> + <i>Trichobius longipes</i>	1	100

Carollia perspicillata (Linnaeus) was infected with seven streblid species.

Prevalence

Nine species of streblids bat fly species showed a prevalence higher than 50% (Table 1) as follows: *Strebla diaemi* Wenzel, 1966 (100%) and *Trichobius diaemi* Wenzel, 1976 (100%) on *Diaemus youngi*; *Trichobius dugesioides* Wenzel, 1966 (100%) on *Lonchorhina aurita* and *Trachops cirrhosus*; *Trichobius joblingi* Wenzel, 1966 (100%) on *Micronycteris minuta* and on *C. perspicillata* (91%); *Megistopoda proxima* (Seguy, 1926) (100%) on *Sturnira lilium*; *Trichobius longipes* Rudow, 1871 (83%) on *P. hastatus*; *Trichobius uniformis* Curran, 1935 (81%) on *Lonchophylla dekeyseri*; *Strebla chrotopteri* Wenzel, 1976 (75%) on *C. auritus* and *Paratrachobius longicrus* (Miranda-Ribeiro, 1907) (67%) on *Platyrrhinus lineatus*.

Host specificity

Abundance of streblid flies varied significantly among their bat hosts ($F_{25} = 206.9$, $p < 0.0001$) (Fig. 3). The *post*

hoc test results indicated that the differences observed were from the associations of *Speria ambigua* Kessel, 1925, *Strebla guajiro* (García et Casal, 1965), *S. mirabilis* (Waterhouse, 1879), *Strebla wiedemanni* Kolenati, 1856, *Trichobius dugesii* Townsend, 1891 and *T. diaemi* (Table 2).

Some bat fly species were more specific to their hosts than others, occurring in greater abundance on those preferred hosts (Fig. 3). For example, *S. wiedemanni* was found in high abundance ($n = 540$), but only on a single host (*S. wiedemanni* in *D. ecaudata* was considered accidental). Likewise, *Trichobius furmani* Wenzel, 1966 was also found in high abundance, but on two hosts only (*D. ecaudata*, $n = 372$ and *D. rotundus*, $n = 57$).

The bat species with the highest number of component communities of ectoparasites was *G. soricina*, with 44% of associations with three streblid species and also 55% with two streblid species (Table 2). Regarding the ectoparasites, the species that had the highest numbers of non-accidental recorded associations were *Trichobius dugesii*, *T. uniformis* and *Strebla mirabilis*, which were found on four host species.

DISCUSSION

The richness of bat flies found (15) was quite similar to the ones found at other sites in the same biome and at the same latitudes. For example, Eriksson et al. (2011) reported 17 species of bat flies in Serra da Bodoquena Park in the state of Mato Grosso do Sul, Graciolli et al. (2010) reported 21 species in Serranópolis in the state of Goiás, whereas Komeno and Linhares (1999) recorded 12 species in Uberlândia in the state of Minas Gerais. Nevertheless, the composition was quite different.

The composition of assemblage of streblid flies on bats from the Cerrado can be considered to be richer than expected. A total of 47 bat fly species have been reported in association with the 26 bat species in the Cerrado (Komeno and Linhares 1999, Graciolli et al. 2006, 2010, Aguiar and Antonini 2011, Eriksson et al. 2011, Santos et al. 2013). According to Santos et al. (2013), factors that may contribute to the high number of bat fly species in the Cerrado are external to the host and can affect the abundance of ectoparasites and their bats, favouring the sampling of host species with different component communities.

Additionally, it is possible to find bat fly species that are of low prevalence on their host species, such as those reported in deciduous forests (Camilotti et al. 2010) and the Caatinga (Rios et al. 2008), who presented a less rich and abundant fauna. Santos et al. (2013) listed were the vegetation structure, the abundance of host shelter and the high number of individuals and host species among the most important factors. Changes in the component communities of ectoparasitic flies on host species can also be attributed to regional differences in the species composition of bats, the biogeographic history of the area, as well as the lack of specificity of ectoparasites (Rui and Graciolli 2005).

Nonetheless, we cannot discard the possibility that trapping bats immediately outside caves at the emergence of evening with simultaneous bat captures of two or more bats may allow for the transference of flies from one host species to another. This contamination, while in the net before transference to individual cloth bags, could also account for the higher number of bat fly species found on bats in this study and the lower host specificity of the flies.

Only three accidental associations were recorded: *Paraeuctenodes longipes* Pessoa et Guimaraes, 1937 and *T. uniformis* on *C. perspicillata* and *S. wiedemanni* on *D. ecaudata*. Excluding these associations, we found the proportion of monoxenous species (56%) to be lower than the 87% found by Dick and Gettinger (2005) and the 88% found by Santos et al. (2013). In contrast, we found a higher proportion of polyxenous species (44%) compared to the proportions found in other studies. For example, Santos et al. (2013) found only one polyxenous species. Some of those species, however, presented a higher level of specificity to a few bat hosts.

This specificity appears to be very uncommon because bat species harbouring more than one fly species seem to be very common in the Neotropics and are reported in many areas of tropical forests (see Wenzel 1976, Bertola et al. 2005, Aguiar and Antonini 2011, Santos et al. 2013). However, as multiple bat species often roost in close as-

sociation in caves, newly emerging flies may migrate to atypical hosts, widening the host range documented here for streblids. Therefore, we cannot be certain if we captured bats at the moment they were infected with newly emerging flies.

The taxonomic relationships of bat flies on *Desmodus rotundus*, *Diaemus youngi* and *Diphylla ecaudata* discussed here are similar to those reported by Aguiar and Antonini (2011) and are richer than those found in other regions of the Cerrado (Graciolli et al. 2010). A novelty in our study is that we found *T. diaemi* and *S. diaemi* on *D. youngi* and *Trichobius furmani* on *D. rotundus*. Graciolli et al. (2010) and Eriksson et al. (2011) found *T. parasiticus* Gervais, 1844 and also found only *S. wiedemanni* individuals on *D. rotundus*. Therefore, we have a new host record for these streblid flies in the Cerrado.

On the bat *Lonchophylla dekeyseri*, five non-transitory bat fly species were found. This is the first report of four bat fly species for *L. dekeyseri* in the Cerrado. Another paper describing bat flies on this host was that of Graciolli and Coelho (2001), who found only *Trichobius lonchophyllae* (Wenzel, 1966). It is worthwhile to stress that the data from Graciolli and Coelho (2001) originated from the same caves that we sampled ten years ago.

The *C. perspicillata* assemblage of bat fly richness in APA da Cafuringa was similar to what was found by Eriksson et al. (2011) in Serra da Bodoquena, Mato Grosso do Sul. We found that *T. dugesii*, *T. tiptoni* Wenzel, 1976 and *T. uniformis* are other non-transitory streblids that parasitise *C. perspicillata* in Brazil. In other localities within the Cerrado, *C. perspicillata* appears to be infected mainly with *T. joblingi* (Komeno and Linhares 1999, Eriksson et al. 2011, Santos et al. 2013). In reality, *T. dugesii* was first reported on *G. soricina* (see Graciolli et al. 2010, Eriksson et al. 2011), *T. tiptoni* on *A. caudifer* (Geoffroy Saint-Hilaire) (Komeno and Linhares 1999, Graciolli et al. 2010, Eriksson et al. 2011) and *T. uniformis* were found on *Glossophaga soricina* by Eriksson et al. (2011) and Graciolli and Dick (2012) and on *D. ecaudata* by Aguiar and Antonini (2011).

Whereas eight bat fly species were previously recorded on *G. soricina*, we found ten bat fly species. When compared with other studies that reported seven species (Eriksson et al. 2011), we found only three bat flies in common with our work. The non-transitory streblids recorded on *G. soricina* in our study site, *Paraeuctenode longipes*, *Strebla guajiro* and *Paratrichobius longicrus* (Miranda-Ribeiro, 1907), were not found in other localities of the Cerrado, indicating that they are new records. However, *Trichobius dugesii* and *Speiseria ambigua* are the same non-transitory species found in other localities in the Cerrado region (Graciolli and Coelho 2001, Graciolli et al. 2010).

Among the five non-vampire bat species that had more than three bat fly species, we found differences in the prevalence and intensity of infection, not only inside the assemblage but also in comparison with other areas within the Cerrado. On *A. caudifer*, a similar prevalence was found between the five species of bat fly (approximately 33%) while Eriksson et al. (2011) found a prevalence of 20% for *T. tiptoni*.

For many bat fly species, a high prevalence was found, which was similar to the results obtained from other localities in the Cerrado. *Strebla chropteri*, for example, was the most prevalent on *C. auritus* (75%), which is similar to the value found by Eriksson et al. (2011).

The prevalence of *T. joblingi* on *C. perspicillata* was higher (91%) than those found in other regions of the Cerrado. A prevalence of 66% was found in Uberlândia (Komeno and Linhares 1999), whereas it was 41% in Serra da Bodoquena (Eriksson et al. 2011) and 55% in Barreirinhas (Santos et al. 2013). A difference in prevalence from the associations between *T. longipes* and *P. hastatus* was also found in our study (83%) in comparison with the 33% recorded by Santos et al. (2013) in northeastern Brazil. The same trend was observed for *P. longicrus* and *P. lineatus* (67%) compared to the 13% reported by Eriksson et al. (2011) in the Cerrado.

In our studied site, the prevalence of *Megistopoda proxima* (Seguy, 1926) on *S. lilium* (100%) was higher than that found in other sites of the Cerrado region (Komeno and Linhares 1999, Eriksson et al. 2011). Thus, the prevalence of *M. proxima* on *S. lilium* appears to be more variable throughout the Atlantic Forest region than in the Cerrado region. This is not in accordance with the findings of Eriksson et al. (2011), who reported a similar per bat pattern of infection on *S. lilium* between sites in the Cerrado and Atlantic Forest regions.

The prevalence of streblids on *G. soricina* is usually low (Graciolli and Rui 2001, Bertola et al. 2005), similar to what we found in APA da Cafuringa (<10%), except for *P. longipes*, *S. guajiro* and *T. joblingi*. According to Santos et al. (2013), this variation in values, observed from the study of different regions, supports the hypothesis that the distribution of streblids in different host populations is highly variable.

Overall comparisons of the available data suggest that the component community of streblids varies more be-

tween the Cerrado and the Atlantic Forest phytogeographical regions than between sites within the same phytogeographical region. Therefore, environmental conditions play a vital role in local bat fly associations, which leads to a variable component community across a wide distribution of host species (Wenzel and Tipton 1966). Prevalence patterns vary extensively between sites, but there is no apparent pattern associated with phytogeographical domains. The local host abundance and shelter type are known to affect the prevalence of streblids (Arneberg et al. 1998).

In conclusion, bats roosting in caves are known to have both higher prevalence and intensity of infection relative to those roosting in more ephemeral situations (Patterson et al. 2007). Therefore, the data available on streblids support the hypothesis that conspecific bats use variable shelters throughout their distributional range and/or that bat species vary widely in their abundance within a phytogeographical region. However, the intensity of infection by streblids on a host species does not vary as much between sites and even between different phytogeographical regions. In the bats sampled by mist nets away from the roost, the generally higher prevalence and intensity of infection could result from the sampling of cave-inhabiting bats, rather than from their conspecific populations roosting in tree holes, leaves or other roost structures. Thus, additional studies at multiple sites are needed to verify the possible patterns of bat fly associations discussed in this study.

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