A new coccidian parasite of the boodie, *Bettongia lesueur* (Mammalia: Marsupialia: Potoroidae), from Australia

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Abstract: Four of 28 wild boodies or burrowing bettongs, *Bettongia lesueur* (Quoy et Gaimard) passed oocysts of species of *Eimeria* Schneider, 1875. The boodies are surviving on off-shore islands and in large predator-proof sanctuaries on the mainland where they were reintroduced. The boodie is a potoroid marsupial extinct from the mainland of Australia due to predation from red foxes and feral cats. Comparison with other species of the genus *Eimeria* indicates that the coccidium found represents a new species. Sporulated oocyst of *Eimeria burdi* sp. n. are pyriform, 21.0–24.0 µm (mean 22.6 µm) by 14.0–16.0 µm (14.9 µm), with a length/width ratio 1.31–1.71 (1.52) and 1-µm-thick yellowish bilayered wall. Micropyle is present at the thinner apex end filled with hyaline body. Polar granules are absent. Sporocysts are ellipsoidal, 10.0–13.5 µm (11.8 µm) by 7.0–8.5 µm (7.4 µm), shape index is 1.42–1.89 (1.63) and a very thin, poorly defined unilayered sporocyst wall is 0.2 µm thick with a domelike almost indistinct Stieda body. Substieda body is indistinct.

Keywords: Coccidia, Apicomplexa, *Eimeria*, morphology, taxonomy, marsupials, bettongs, new species

Bettongs, potoroos and two of the rat-kangaroos belong to family Potoroidae Gray, a group of rabbit-sized marsupials endemic to Australia. The family includes four genera: *Aepyprymnus* Garrod, *Bettongia* Grey, *Caloprymnus* Thomas and *Potorous* Desmarest. Prior to European settlement there were 12 potoroids. In the past two centuries, four species have become extinct and two survive only on off-shore islands, despite previous distribution on the mainland (Woinarski et al. 2014).

The boodie or burrowing bettong, *Bettongia lesueur* (Quoy et Gaimard) (Mammalia: Marsupialia: Potoroidae) was once common in its former distribution across a wide span of the southern and central mainland of Australia. It is now extinct from the mainland due to predation from red foxes, *Vulpes vulpes* (Linnaeus), and feral cats, *Felis catus* Linnaeus, and survives only on a handful of small islands off the west coast of Australia and in large predator-proof sanctuaries on the mainland (Australian Wildlife Conservancy [AWC]’s Scotia and Yookamurra Sanctuaries; Arid Recovery) into which boodies have been reintroduced (Short and Turner 2000).

Parasitological studies on potoroids are scarce and for the boodie non-existent. Recent studies have focused on blood parasites, *Trypanosoma* spp., as a potential contributing factor in potoroids decline (Botero et al. 2013, Thompson et al. 2014). There are no coccidian parasites described from the boodie, despite four species of *Eimeria* Schneider, 1875 described from other potoroids (Barker et al. 1988, Duszynski 2016a). Here, we describe a new species of *Eimeria* from *B. lesueur* and compare it to those from potoroids.

MATERIALS AND METHODS

Faecal samples (n = 28) from 28 individual *Bettongia lesueur*, the burrowing bettong or boodie, were collected in April, 2016 from animals living in AWC’s Scotia Sanctuary, New South Wales 2648, Australia (33°09’49”S; 141°07’38”E; Geodetic Coordinate System Australia 1994). This reintroduced population of *B. lesueur* lives wild in one of two 4000 ha predator-proof fenced areas within the sanctuary.

Samples were collected from either the trap or bag holding the animal or directly from the animal and a portion placed directly into a sodium-acetic acid formalin (SAF) solution and held at room temperature. Routine faecal flotation on fresh samples was performed to reveal presence of detectable parasitic stages. Faecal samples preserved in SAF were received at the Faculty of Veterinary Science, The University of Sydney. Sporulated oocysts were examined and photographed using Nomarski interference contrast microscopy. Thirty sporulated oocysts were measured using a calibrated ocular micrometre. Measurements in micrometres are reported as range followed by mean in parentheses.

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RESULTS

_Eimeria burdi_ sp. n.  
Figs. 1–7

**ZooBank number for species:** urn:lsid:zoobank.org:act:CD9EB84E-6D53-4225-B374-60D66EFDA75D

**Description** (based on 30 sporulated oocysts): Sporulated oocysts pyriform, 21.0–24.0 (22.6) by 14.0–16.0 (14.9); shape index (length/width) 1.31–1.71 (1.52). Oocyst wall smooth, pale yellow. Oocyst wall bilayered, 1 thick (outer layer 0.8, inner layer 0.2). Oocyst residuum and polar granule absent. Thinner apex end with micropyle, approximately 4 across. Hyaline body filling apex of oocyst, mostly indistinct. Sporocysts ellipsoidal, 10.0–13.5 (11.8) by 7.0–8.5 (7.4), with very thin, smooth, poorly defined unilayered sporocyst wall 0.2 thick. Sporocyst length/width ratio 1.42–1.89 (1.63). Stieda body almost indistinct, domelike, ~1 × 0.5. Substieda body indistinct. Sporocyst residuum present, composed of numerous granules of approximately 0.2–0.5 each, condensed into oval to irregular clusters of approximately 5 in diameter, filling space between sporozoites. Each sporocyst contains two sporozoites. Sporozoites broadly elongate, arranged head to tail within sporocyst. Sporozoites with two distinct refractile bodies, a larger (2–3) and a smaller (1–2). Sporozoite nucleus oval but indistinct, situated between refractile bodies. Sporozoites and sporozoite residuum float free within sporocyst. Sporozoite residuum not enclosed in membrane.

**Type and only known host:** _Bettongia lesueur_ (Quoy et Gaimard) (Mammalia: Marsupialia: Potoroidae), burrowing bettong or boodie.

**Type and only known locality:** 33°09′49″S; 141°07′38″E; AWC Scotia Sanctuary, via Wentworth, New South Wales 2648, Australia.

**Distribution:** Unknown.

**Sporulation:** Sporulated oocysts recovered from faeces transported from locality to the laboratory. The sample spent varying periods in transit (less than 10 hours) before reaching the laboratory where it was stored in SFA.

**Prevalence:** 14% (found in four out of 28 examined faecal animals).

**Prepatent and patent periods:** Unknown.

**Site of infection:** Unknown, oocysts recovered from faeces.

**Endogenous stages:** Unknown.

**Type specimens:** Phototypes are deposited at the Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, Branišovská 31, České Budějovice, Czech Republic (No. IP Prot Coll 36).

**Etymology:** The specific epithet burdi reflects the host local name ‘burdi’ of the Nyungar people of Western Australia and is given, in accordance with the International Code of Zoological Nomenclature (Article 51.1), as a noun in apposition (ICZN 1999).

**Remarks.** To these date, there are four species of coccidian parasites described from Potoroidae (Barker et al. 1988, Duszynski 2016a). Oocysts of our new species of _Eimeria_ are different from those of all previously described species by size and shape, presence of a micropyle and absence of polar granule and oocyst residuum. The defined pyriform shape, smooth wall and size of oocysts, 21.0–24.0 µm (22.6 µm) by 14.0–16.0 µm (14.9 µm), are the most distinct characters of _Eimeria burdi_ sp. n. The closest oocyst of _Eimeria potoroi_ Barker, O’Callaghan et Beveridge, 1988 from _Potorus tridactylus_ (Kerr) (long-nosed potoroo) is 26.2 µm × 18.5 µm and was reported to be slightly pyriform and its wall radially striated with mamillated surface.

Oocysts of _Eimeria gaimardi_ Barker, O’Callaghan et Beveridge, 1988 from _Bettongia gaimardi_ (Desmarest) (Tasmanian bettong) are also slightly pyriform with mamillated surface. Oocysts of _Eimeria aepyprymni_ Barker, O’Callaghan et Beveridge, 1988 from _Aepyprymnus rufescens_ (Gray) (rufous bettong) and _E. gaimardi_ are larger, 36.7 µm × 21.9 µm and 34.6 µm × 24.3 µm, respectively, than those of _Eimeria burdi_. _Eimeria mundayi_ Barker, O’Callaghan et Beveridge, 1988 from the long-nosed potoroo are smaller, spherical and possessing a polar granule (Barker et al. 1988, Duszynski 2016a).

**DISCUSSION**

Many coccidian parasites are host-specific (Barker et al. 1988, Duszynski 2016a) and the effect of these parasites on health and disease of potoroids is unknown. Reintroductions of declined animal populations should take into account the role of infectious agents such as parasites in the disease risk assessment (Jakob-Hoff et al. 2014). Parasites have to be considered when preserving host species from extinction, because with the host extinction, their microbiota, including their unique parasites, are also lost.
forever (Daszak et al. 2011). Coccidiosis (disease caused by species of *Eimeria*) can potentially be life-threatening in young captive marsupials (Vogelnest and Portas 2008).

Reports on coccidian parasites from potoroids are so far restricted to detection of oocysts in faeces from healthy wild or captive animals (Barker et al. 1988; the present study). For example, year-long presence of large numbers of *Eimeria echidnae* Barker, Beveridge et Munday, 1985 in long-term captive short-beaked echidnas, *Tachyglossus aculeatus* (Shaw), has not been associated with any health issues (Debenham et al. 2012). In fact, *E. echidnae* are considered normal microbiota of captive echidnas without the need for therapeutic management.

Seasonal occurrence of *Eimeria bardi* sp. n. across the very restricted distribution of the boodie is required to better understand the role the parasites play in health and disease. Similarly, it remains to be shown whether boodies from the Western Australian island whence the reintroduced animals were sourced are infected with *E. bardi*.

Recently, it has been argued that parasitologists have examined only 13% of all vertebrate species for coccidia (Duszynski 2016b). It is expected that a large majority of coccidia in Australia are unknown and yet to be described and their role in health and disease is yet to be understood.

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REFERENCES


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